



REVIEW ARTICLE

An association of metabolic syndrome constellation with cellular membrane caveolae

Wei-zheng Zhang*

CMP Laboratory, Port Melbourne, Melbourne, Victoria, Australia

Metabolic syndrome (MetS) is a cluster of metabolic abnormalities that can predispose an individual to a greater risk of developing type-2 diabetes and cardiovascular diseases. The cluster includes abdominal obesity, dyslipidemia, hypertension, and hyperglycemia – all of which are risk factors to public health. While searching for a link among the aforementioned malaises, clues have been focused on the cell membrane domain caveolae, wherein the MetS-associated active molecules are colocalized and interacted with to carry out designated biological activities. Caveola disarray could induce all of those individual metabolic abnormalities to be present in animal models and humans, providing a new target for therapeutic strategy in the management of MetS.

Keywords: *metabolic syndrome; caveolae; dyslipidemia; hypertension; hyperglycemia; caveolins*

*Correspondence to: Wei-zheng Zhang, 86 Bay Street, Port Melbourne, Vic. 3207, Australia, Email: weizzhang@hotmail.com

Received: 3 January 2014; Revised: 21 January 2014; Accepted: 21 January 2014; Published: 12 February 2014

Metabolic syndrome (MetS) is defined as a cluster of metabolic abnormalities which is predictive of a high risk of type-2 diabetes and cardiovascular disease (1,2). Despite slight differences in its definition, the major clinical features encountered are insulin resistance (IR), hyperglycemia, obesity or abdominal obesity, and hypertension and dyslipidemia (3). Although there have been many proposals on the pathogenesis and pathophysiology, any possible mechanism to connect all such abnormalities at cellular and molecular levels has not been delineated. Gathering from available literature, it becomes plausible that the cell membrane domain, caveolae, plays a vital role in the formation of MetS.

The cell bilayer membrane serves as a barrier to the surroundings. Caveola is a special type of lipid raft, which appears as a minute invagination (50–100 nanometer) on the membrane in many vertebrate cell types, especially in endothelial cells (EC) and adipocytes (4–6). These flask-shaped structures are rich in proteins and lipids, and selectively perform an important role in the exchange of information and materials with the environment by acting as coordinators of signal transduction and endocytosis. It has been well documented that caveolae act as signaling platforms, function as a gathering point for numerous signaling molecules, manage cell volume and volume-sensitive signaling in an adipocyte system, act as mechanosensors in many cell types, and control fluctua-

tion through many well-defined signaling cascades (7–9). Some pathogens can enter the target cells via caveolae to preclude a self-degradation in lysosomes (8,9). The formation of caveolar endocytic vesicles is dependent on the ability of caveolin oligomerization, and sufficient levels of cholesterol and caveolin insertion in plasma membrane (10). Many active transmembrane proteins, including transporters, receptors, enzymes and caveolins, are colocalized within caveolae. They have a distinctive constitution with highly enriched cholesterol, sphingolipids, and 1-, 2-, and 3-caveolins as coating proteins (11). Munday and colleagues have demonstrated a caveolae involved in multi-step trafficking pathway of endocytosis (9), whereby caveolin-containing bodies ‘cavicles’ are transported from the ‘caveosome’ to the cell surface caveolae via microtubules (8,9).

A new general function of caveola has been revealed by its role in the uptake of different metabolites of lipid and glucose metabolism (12). Caveolae, therefore, come into sight as important centers for many facets of metabolism serving as gateways for the uptake of nutrients across the cell membrane, and as platforms for the metabolic conversion of nutrients, especially in adipocytes and ECs (12). Caveolins, the integral plasma membrane proteins present in caveolae with proven scaffolding, transport, and signaling capabilities, have emerged as key players in lipid dynamics and membrane microdomain disorders apart

from its role in the obesity and IR development (13). Currently available literature on caveola and its functions may allude to our understanding of obesity, diabetes, and other metabolic disorders. However, so far there are no definitive published studies on the linkage of MetS and caveolae to raise attention to a mechanism of MetS formation, which have great potential for MetS therapeutic management.

Herein, the aim of this opinion review is to provide an overview from available literature on the participation of caveolae and caveolins as common novel modulators in pathogenesis of the MetS cluster.

The active molecules involved in the MetS cluster are associated with membrane caveolae.

Insulin resistance

Membrane caveolae play a vital role in compartmentalization of insulin signaling (14). A primary basis of type-2 diabetes is IR caused by a dysfunction of insulin signaling in target tissues. Under the pathological condition of IR, the body produces insulin but cells fail to respond to the normal actions of the insulin hormone due to any efficiency changes of insulin surface receptors. The insulin receptor and part of the downstream signaling mediators can be recruited to and gather at caveolae (15). As part of the signaling, the auto-phosphorylated insulin receptor in primary adipocytes promptly engages with tyrosine phosphorylation of caveolin-1 upon being internalized through a caveolae-mediated process (16), and the process requires membrane cholesterol (17). Lack of caveolae induces IR in animals and human beings (15). The efficacy of insulin signaling in the adipocyte can be firmly relied on the localization of at least two insulin-responsive elements (insulin receptor and GLUT4) to caveolae as well as on a direct functional interaction between insulin receptor and caveolin-1 (18). Approximately 19% of insulin receptor molecules were detected in caveolar regions of adipocytes, and an aberrant ganglioside distribution in the caveolae constitution resulted in IR (19). Disarrays in caveolae lipid composition have been shown *in vitro* to extricate proteins from caveolae, thereby altering their normal functionality and the consequent downstream signaling (20). Furthermore, a high-cholesterol diet can induce an enhanced insulin-induced insulin receptor activation, impair the downstream molecules IRS-1 and Akt activities, and abolish an induction of caveolin-1 tyrosine phosphorylation under the insulin stimulation (20). The atypical interaction between insulin receptor and gangliosides within caveolae can also be one of the molecular pathogeneses of type-2 diabetes (21).

Given that cell volume regulation is significantly adjusted upon adipocyte differentiation which is associated with the number of caveolae present on the cell surface, it is insinuated that caveolae are involved in the pathogenetic mechanism of IR and MetS (22). Although

the responses to insulin may vary among cell types, caveolae engage in the process. In RLE-6TN cells, insulin is taken up through endocytosis accompanied by the insulin receptor within caveolae (23). Concomitantly, insulin uptake in EC requires expression of caveolin-1, the main coating protein of caveolae, supporting the motion of caveolae mediating insulin uptake (24). In the adipose cell, caveolin-1 and caveolae also regulate insulin action. Loss function of caveolin-1 reduces maximal insulin response through a lowered stability and diminished expression of the insulin receptor and GLUT4 (25). In the liver cells, caveolin greatly enhances the insulin receptor signaling upon being over-expressed *in vivo* (20), acts as an important regulator of glucose metabolism that can augment insulin signals in the obese mouse livers attributing mostly to an increased insulin receptor activity and the caveolin-mediated direct inhibition of protein tyrosine phosphatase 1B (26). Caveolae and caveolin-1 also play a role in insulin-like growth factor-I receptor internalization and function modulation, respectively (27). A high-cholesterol diet alters caveolin-1 expression *in vivo*, and insulin receptor localization as well as activity (20), which may imply a linkage of hyperlipidemia and IR via caveolae. Strong evidence authenticates that caveolae participate in the pathological origin of IR.

Hyperglycemia

The major glucose transporter GLUT4 manipulates the blood glucose level and its cellular localization within caveolae is strictly a prerequisite for the functionality of cellular glucose transportation and maintenance (28). Other glucose transporters, GLUT1 and GLUT3 together with hexokinase (a glycolytic and glycogenic priming enzyme) are also found in caveolae (29). In skeletal muscle and adipose tissue, GLUT4 is translocated, upon insulin stimulation, from intracellular storage compartments to cellular membrane within caveolae to trigger the insulin-stimulated glucose uptake, and the process of GLUT4 protein cellular trafficking is precisely regulated by the insulin receptor signals through a series of highly organized membrane trafficking events (30–32). Elevated glucose concentrations diminish the number and the size of caveolae (33), and cause disarray of caveolae constitution (34). In the diabetic lung, however, the EC displays an increased number of caveolae, an enlarged surface area, escalated cholesterol content, and an over-expression (gene and protein) of caveolin-1 (35). Certainly a compensational mechanism needs to be ascertained. The study on the caveolin-1 knockdown adipocytes demonstrated reductions in the insulin-triggered GLUT4 recruitment to the cell surface, in the insulin-stimulated glucose transport, insulin receptor activation caused by a reduced stability, and in expressions of insulin receptors and GLUT4 (25), reversely confirming the roles of caveolae in glucose transportation and metabolism. Further *in vitro*

studies also demonstrated that a depletion of cholesterol contained in caveolae can partially inhibit GLUT4 internalization in L6 myoblasts (36), and that IR can be bypassed by manipulating GLUT4 endocytosis for maintaining sufficient GLUT4 on the surface (37). Caveolae formation requires cavin. The cavin-knockout mice are viable and of normal weight but illustrate a significant glucose intolerance, hyperglycemia, and hyperinsulinemia (18). Concisely, glucose transportation and metabolism require its major transporter GLUT4 to exist on the cell surface within caveolae. Any disruption of caveolae could result in hyperglycemia.

Obesity and dyslipidemia

Obesity, especially visceral obesity, causes IR and is associated with dyslipidemia, impaired glucose metabolism, and hypertension (38). Dyslipidemia commonly accompanies obesity and MetS (39). Caveolae physically involve in fatty acid (FA; 40,41), triacylglycerol (42), and cholesterol (43) uptakes, in the protection of adipocyte from the lipotoxic effects of elevated FAs, and in the cholesterol synthesis (44). Caveolae within adipocyte are regulated by energy homeostasis. At a state of positive energy balance, expressions of some proteins are accordingly increased to meet the requirements of caveolae structure remoulding coping with the adipose tissue expansion for maximizing the capacity of cellular lipid storage (45). Rising circulating FA levels can also induce a substantial upregulation of caveolar proteins (45). In the obese rats, however, caveolae are not evenly transformed. In the endothelium of arteries, the number of caveolae is declined, whereas, at the ends of smooth muscle cells, caveolae density is intensified (46), which may be aligned with the pathophysiology of obesity on vascular function and carbohydrate metabolism. It also has been demonstrated that lacking functional caveolae can cause dyslipidemia and can result in reduced fat storage accompanied by smaller sizes of the fat cells in mice and humans (45). The cellular FA utilization and the initial metabolic regulation require a precise control on the cellular FAs uptake via membrane. In many tissues, caveola facilitates a major fraction of FA uptake (47) and conversely higher levels of FAs can induce an enhanced density of caveolae in obese rats (48). A study on dietary intake of saturated fat implies that the saturated fats can intensify the levels of sphingolipids in cardiac cell membranes resulting in a disruption of the caveolae composition and can significantly reduce the systolic contractile performance as well as caveolin-1 contents (49). Investigation on cavin-knockout mice, which has a disrupted caveolae composition, demonstrated an extensively shrunken adipose tissue mass (18). The membrane lipid environment affects caveolin isoforms (49). In the arterioles of obese rats, only caveolin-1 and -2 oligomer expressions, but not their monomers and caveolin-3,

are elevated (46). Sphingomyelin is one of the major phospholipids of caveolae. Conformational changes in plasma membrane sphingomyelin can induce obesity and type-2 diabetes (50). In the caveolin-1 null mice, triglyceride and free FA levels are significantly elevated (51), and a modestly increased rate of lipolysis together with a diminished cellular integrity are evidenced (45). In contrast, caveolin-1-knockout mice exhibit a lean phenotype with an overt resistance to the diet-induced obesity, whereas, caveolin-3-knockout mice show a marked IR accompanied by an increased body weight and adiposity in a normal diet (13).

Cluster of Differentiation 36 (CD36), a member of the class B scavenger receptor and FA transporter family within the caveolae domain on cell surface, binds many ligands including oxidized low-density lipoprotein (OxLDL) (52), native lipoproteins (53), oxidized phospholipids (54), and long-chain FAs (55). Together with Caveolin-1, CD36 participates in adipocyte FA uptake and metabolism, and both are coordinately involved in lipid droplet formation (50) and associated with long-chain FA uptake (56). CD36 membrane levels and the turnover are abnormal in diabetes, causing a dysfunctional FA utilization. In addition, polymorphism of the CD36 gene has been shown to influence its susceptibility to MetS (47). OxLDL impacts on membrane rafts and caveolae in the distribution of different membrane raft constituents, on membrane cholesterol composition, and on lipid packing of different membrane domains (57). OxLDL moves CD36 from low to high buoyant density membrane fractions together with caveolin-1 (58). Upon uptake OxLDL, CD36 can segregate them at the cell surface, interfering with intracellular trafficking and degradation (59). Conversely, Caveolin-1 plays an important role in recruiting fatty acid translocase (FAT)/CD36 to caveolae, and in regulating FA uptake via cellular surface availability of FAT/CD36 (60). Caveolae integrity has been shown to directly affect the CD36 function and blood pressure regulation (60). Both high-density lipoprotein cholesterol (HDL) and low-density lipoprotein cholesterol (LDL) participate in maintaining the lipid environment in caveolae (61,62). The former upholds the caveolae integrity, while the latter, especially oxLDL, acts on an opposed effect of inducing membrane caveolae internalization (62).

A large amount of evidence suggests that the abnormality of lipids could affect caveolae composition and functionality, which would further disturb FA metabolism in the development of dyslipidemia and obesity.

Hypertension

Although many abnormalities are suggested to be the pathogenesis of hypertension, endothelial nitric oxide synthase (eNOS) existing within cell membrane caveolae, which produces nitric oxide (NO), is well defined as a major factor in maintaining vascular function. A reduced

production of NO can cause hypertension (62–64). An aberrant regulation on eNOS together with an associated NO/SNO release is directly linked with various vascular diseases. eNOS is primarily localized on the Golgi apparatus and plasma membrane caveolae in EC (64). It functions to produce NO only when it presents on the cell surface within caveolae (62). Caveolae transduce eNOS/NO/SNO cardioprotective signaling in the heart (65). The scaffolding domain of caveolin-1 serves as an endogenous negative regulator of eNOS function (66), which is supported by the evidence of that the caveolin-deficient animals show an unopposed NO production promoting vessel dilation (67). In addition, reactive-oxygen-species-dependent eNOS activation and eNOS uncoupling are critically regulated by caveolin-1 (67). Alpha-galactosidase A participates in glycolipid metabolism and caveolin assembly. Research on alpha-galactosidase A-knockout mice has demonstrated that the animals had high molecular weight caveolin oligomers reduced and detached caveolae in the ECs (68).

Caveolae also associate functionally and anatomically with many other active molecules of blood pressure ‘regulators’. Specifically, caveolae act as binding sites for calcium ions (as a major Ca(2+)), influx is through L-type calcium channels within caveolae signaling domains) (69), participate in regulating both pumping and signal transducing functions of Na(+)/K(+)-ATPase (70), are involved in AT1receptor internalization (71), influence calcium-activated chloride channel properties (72), and regulate alpha1-adrenergic receptor signaling (73). In addition, caveolae engage critically in endothelial signal transduction from shear stress to vasodilator production and release (74). The endothelium-dependent shear-stress-mediated vasodilation requires the integrity of caveolae. In the Cav-1^{-/-} endothelium, the shear-stress-mediated vasodilators including NO, epoxyeicosatrienoic acids, and prostaglandins are deficient (74). In view of that, caveolae and caveolin participate in the regulation of blood pressure. Deformity of caveolae can cause hypertension.

Some other caveolae associated active molecules involved in MetS

Many other active molecules within caveolae have also been revealed to be associated with MetS. Vitamin D (25(OH)D) deficiency has been demonstrated to be associated with a high risk of MetS involving in a higher serum triglyceride, elevated fasting glucose, and induced IR (75). Vitamin D initiates rapid cellular responses through a putative plasma membrane-associated vitamin D receptor (VDR), which is sited on caveolae (Pdia3) (76). VDR regulates gene expression of the encoding proteins that propagate the traditional genomic functions of vitamin D, and the quick response of vitamin D binding with VDR delays chronic diseases of aging such as cancer,

vascular disease, type-1 and -2 diabetes, arteriosclerosis, osteoporosis, and infection (77). Binding with VDR at different cellular locations can selectively mediate both genomic and cellular responses (78). VDR can be recruited on cell surface within caveolae upon stimulation of vitamin D (79). Elevated 7-dehydrocholesterol (a cholesterol precursor which is converted to vitamin D3 in the skin) in the caveolar membrane can induce a defective caveolar signaling (80). As its roles in metabolism become evident, a larger vitamin D clinical trial has been recently approved by NIH on its prevention of diabetics (news release: <http://www.nih.gov/news/health/oct2013/niddk-21.htm>), so the final results of any promises of therapeutic benefits beyond healthy bones are pending.

The beneficial metabolic actions of estrogen-based therapies are mainly mediated by estrogen receptor (ER). ER has been suggested to be involved in obesity (81). Upon binding with ER at caveolae, estrogen can induce NOS activation (61) and play a role in the maintenances of body metabolism (82). The ER knockout-mouse electively developed an accelerated weight gain, massive adiposity, severe IR, and glucose intolerance (83). 17β-estradiol administration regulates some key metabolic genes in insulin-sensitive tissues and confers a strong protection against high-fat diet-induced metabolic disturbances (83). Despite the beneficial estrogenic effects in reversing some of the MetS symptoms, an increasing body of evidence now links estrogenic signaling with MetS (84). Targeted estrogen delivery on ER reverses MetS (85). Specifically, loss of ER signaling leads to IR and obesity in animals (86).

Inflammation can also be involved in caveolae impinging on MetS. It has been suggested that NF-κB activation may participate in chronic inflammation, IR, endothelial dysfunction, hypertension, and dyslipidemia (87). NF-κB activation, however, is associated with caveolae and specifically requires caveolin-1 (88). Moreover, during the inflammation process and under hyperglycemia, the apoptosis of macrophages might occur, leading to the spreading of lipids of cell membrane caveolae from macrophages into intracellular spaces in the vessel wall (33). The damaging vascular wall can insert detrimental effects on the development of all relevant abnormalities of MetS. Furthermore, exercise could reduce MetS and exert anti-inflammatory effects (89), which may act through increasing shear-stress to regulate caveolae signal transduction (74).

Future studies and prospective applications

As the joint AHA-NHLBI statement has recommended MetS as a clinical entity, and for its involvement in all MetS relevant abnormalities (90), caveolae are a potential therapeutic target for MetS. Controlling obesity by reducing hyperlipidemia and LDH (especially oxLDL) level and increasing physical activities, drugs for reducing

blood lipids and/or blood pressure have shown benefits in the management of MetS. At the molecular level, manipulating caveolins/caveolae can affect NO production, insulin action, and lipid and hormone metabolisms. Therefore, caveolae together with its components may become useful targets for treating MetS. For example, L-Arginine supplementation could induce its transporter, cationic amino acid transporter 1, recruited into caveolae and concurrently trigger caveolae translocation onto cell surface (62) with subsequent effects on insulin sensitivity and reduce body fat content (91). Preliminary studies in our laboratory on caveolar cellular internalization have demonstrated a potential regulation on the function of caveolae in EC after treatment with nutrients and chemicals (unpublished data). Extensive research on the regulation of caveolae function, especially on cellular trafficking both *in vivo* and *in vitro*, is warranted. In conclusion, the rising prevalence of the MetS especially in youth (92) and its associated abnormalities is a major public health problem. However, the physiopathology of MetS remains to be ascertained. Information from published studies suggests that all MetS-associated metabolic abnormalities have a common pathogenesis origin emanated from cell membrane caveolae, which presents a new therapeutic strategy for MetS.

Conflict of interest and funding

The author has not received any funding or benefits from industry or elsewhere to conduct this study.

References

- Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. *Diabetes*. 1988; 37(12): 1595–607.
- Expert Panel on Detection, Evaluation, Treatment of High Blood Cholesterol in Adults. Executive summary of the Third Report of The National Cholesterol Education Program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (Adult Treatment Panel III). *JAMA*. 2001; 285(19): 2486–97.
- Onat A. Metabolic syndrome: nature, therapeutic solutions and options. *Expert Opin Pharmacother*. 2011; 12(12): 1887–900. doi: 10.1517/14656566.2011.585462.
- Vinent J, Johnsen AH, Roepstorff P, Harpott J, Tranum-Jensen J. Identification of a major protein on the cytosolic face of caveolae. *Biochim Biophys Acta*. 2005; 1717(1): 34–40. doi: 10.1016/j.bbamem.2005.09.013.
- Liu L, Pilch PF. A critical role of cavin (polymerase I and transcript release factor) in caveolae formation and organization. *J Biol Chem*. 2008; 283(7): 4314–22. doi: 10.1074/jbc.M707890200.
- Hill MM, Bastiani M, Luetterforst R, Kirkham M, Kirkham A, Nixon SJ, et al. PTRF-Cavin, a conserved cytoplasmic protein required for caveola formation and function. *Cell*. 2008; 132(1): 113–24. doi: 10.1016/j.cell.2007.11.042.
- Insel PA, Patel HH. Membrane rafts and caveolae in cardiovascular signaling. *Curr Opin Nephrol Hypertens*. 2009; 18(1): 50–6. doi: 10.1097/MNH.0b013e3283186f82.
- Pelkmans L, Kartenbeck J, Helenius A. Caveolar endocytosis of simian virus 40 reveals a new two-step vesicular-transport pathway to the ER. *Nat Cell Biol*. 2001; 3(5): 473–83. doi: 10.1038/35074539.
- Mundy DI, Machleidt T, Ying YS, Anderson RG, Bloom GS. Dual control of caveolar membrane traffic by microtubules and the actin cytoskeleton. *J Cell Sci*. 2002; 115(Pt 22): 4327–39.
- Jansen M, Pietiainen VM, Polonen H, Rasilainen L, Koivusalo M, Ruotsalainen U, et al. Cholesterol substitution increases the structural heterogeneity of caveolae. *J Biol Chem*. 2008; 283(21): 14610–18. doi: 10.1074/jbc.M710355200.
- Fridolfsson HN, Patel HH. Caveolin and caveolae in age associated cardiovascular disease. *J Geriatr Cardiol*. 2013; 10(1): 66–74. doi: 10.3969/j.issn.1671-5411.2013.01.011.
- Ortegren U, Aboulaiach N, Ost A, Stralfors P. A new role for caveolae as metabolic platforms. *Trends Endocrinol Metab*. 2007; 18(9): 344–9. doi: 10.1016/j.tem.2007.08.007.
- Fruhbeck G, Lopez M, Dieguez C. Role of caveolins in body weight and insulin resistance regulation. *Trends Endocrinol Metab*. 2007; 18(5): 177–82. doi: 10.1016/j.tem.2007.04.001.
- Kabayama K, Sato T, Saito K, Loberto N, Prinetti A, Sonnino S, et al. Dissociation of the insulin receptor and caveolin-1 complex by ganglioside GM3 in the state of insulin resistance. *Proc Natl Acad Sci U S A*. 2007; 104(34): 13678–83. doi: 10.1073/pnas.0703650104.
- Stralfors P. Caveolins and caveolae, roles in insulin signalling and diabetes. *Adv Exp Med Biol*. 2012; 729: 111–26. doi: 10.1007/978-1-4614-1222-9_8.
- Fagerholm S, Ortegren U, Karlsson M, Ruishalme I, Stralfors P. Rapid insulin-dependent endocytosis of the insulin receptor by caveolae in primary adipocytes. *PLoS One*. 2009; 4(6): e5985. doi: 10.1371/journal.pone.0005985.
- Sanchez-Wandeler J, Davalos A, Herrera E, Giera M, Cano S, de la Pena G, et al. Inhibition of cholesterol biosynthesis disrupts lipid raft/caveolae and affects insulin receptor activation in 3T3-L1 preadipocytes. *Biochim Biophys Acta*. 2009; 1788(9): 1731–9. doi: 10.1016/j.bbamem.2009.05.002.
- Liu L, Brown D, McKee M, Lebrasseur NK, Yang D, Albrecht KH, et al. Deletion of Cavin/PTRF causes global loss of caveolae, dyslipidemia, and glucose intolerance. *Cell Metab*. 2008; 8(4): 310–17. doi: 10.1016/j.cmet.2008.07.008.
- Sekimoto J, Kabayama K, Gohara K, Inokuchi J. Dissociation of the insulin receptor from caveolae during TNFalpha-induced insulin resistance and its recovery by D-PDMP. *FEBS Lett*. 2012; 586(2): 191–5. doi: 10.1016/j.febslet.2011.12.019.
- Hahn-Obercyger M, Graeve L, Madar Z. A high-cholesterol diet increases the association between caveolae and insulin receptors in rat liver. *J Lipid Res*. 2009; 50(1): 98–107. doi: 10.1194/jlr.M800441-JLR200.
- Huhtakangas JA, Olivera CJ, Bishop JE, Zanello LP, Norman AW. The vitamin D receptor is present in caveolae-enriched plasma membranes and binds 1 alpha,25(OH)2-vitamin D3 in vivo and in vitro. *Mol Endocrinol*. 2004; 18(11): 2660–71. doi: 10.1210/me.2004-0116.
- Eduardsen K, Larsen SL, Novak I, Lambert IH, Hoffmann EK, Pedersen SF. Cell volume regulation and signaling in 3T3-L1 pre-adipocytes and adipocytes: on the possible roles of caveolae, insulin receptors, FAK and ERK1/2. *Cell Physiol Biochem*. 2011;28(6):1231–46. doi: 10.1159/000335855.
- Oda K, Yumoto R, Nagai J, Katayama H, Takano M. Mechanism underlying insulin uptake in alveolar epithelial cell line RLE-6TN. *Eur J Pharmacol*. 2011; 672(1–3): 62–9. doi: 10.1016/j.ejphar.2011.10.003.
- Wang H, Wang AX, Barrett EJ. Caveolin-1 is required for vascular endothelial insulin uptake. *Am J Physiol Endocrinol Metab*. 2011; 300(1): E134–44. doi: 10.1152/ajpendo.00498.2010.

25. Gonzalez-Munoz E, Lopez-Iglesias C, Calvo M, Palacin M, Zorzano A, Camps M. Caveolin-1 loss of function accelerates glucose transporter 4 and insulin receptor degradation in 3T3-L1 adipocytes. *Endocrinology*. 2009; 150(8): 3493–502. doi: 10.1210/en.2008-1520.
26. Otsu K, Toya Y, Oshikawa J, Kurotani R, Yazawa T, Sato M, et al. Caveolin gene transfer improves glucose metabolism in diabetic mice. *Am J Physiol Cell Physiol*. 2010; 298(3): C450–6. doi: 10.1152/ajpcell.00077.2009.
27. Salani B, Passalacqua M, Maffioli S, Briatore L, Hamoudane M, Contini P, et al. IGF-IR internalizes with Caveolin-1 and PTRF/Cavin in HaCat cells. *PloS One*. 2010; 5(11): e14157. doi: 10.1371/journal.pone.0014157.
28. Cohen AW, Combs TP, Scherer PE, Lisanti MP. Role of caveolin and caveolae in insulin signaling and diabetes. *Am J Physiol Endocrinol Metab*. 2003; 285(6): E1151–60. doi: 10.1152/ajpendo.00324.2003.
29. Rauch MC, Ocampo ME, Bohle J, Amthauer R, Yanez AJ, Rodriguez-Gil JE, et al. Hexose transporters GLUT1 and GLUT3 are colocalized with hexokinase I in caveolae micro-domains of rat spermatogenic cells. *J Cell Physiol*. 2006; 207(2): 397–406. doi: 10.1002/jcp.20582.
30. Karlsson M, Thorn H, Parpal S, Stralfors P, Gustavsson J. Insulin induces translocation of glucose transporter GLUT4 to plasma membrane caveolae in adipocytes. *FASEB J*. 2002; 16(2): 249–51. doi: 10.1096/fj.01-0646fje.
31. Ros-Baro A, Lopez-Iglesias C, Peiro S, Bellido D, Palacin M, Zorzano A, et al. Lipid rafts are required for GLUT4 internalization in adipose cells. *Proc Natl Acad Sci U S A*. 2001; 98(21): 12050–5. doi: 10.1073/pnas.211341698.
32. Kanzaki M. Insulin receptor signals regulating GLUT4 translocation and actin dynamics. *Endocr J*. 2006; 53(3): 267–93.
33. Hayashi T, Juliet PA, Miyazaki A, Ignarro LJ, Iguchi A. High glucose downregulates the number of caveolae in monocytes through oxidative stress from NADPH oxidase: implications for atherosclerosis. *Biochim Biophys Acta*. 2007; 1772(3): 364–72. doi: 10.1016/j.bbadic.2006.11.011.
34. Smart EJ, Li XA. Hyperglycemia: cell death in a cave. *Biochim Biophys Acta*. 2007; 1772(5): 524–6. doi: 10.1016/j.bbadic.2007.01.005.
35. Uyy E, Antohe F, Ivan L, Haraba R, Radu DL, Simionescu M. Upregulation of caveolin-1 expression is associated with structural modifications of endothelial cells in diabetic lung. *Microvas Res*. 2010; 79(2): 154–9. doi: 10.1016/j.mvr.2009.11.008.
36. Engelbrecht B, Stratmann B, Hess C, Tschoepe D, Gawlowski T. Impact of GLO1 knock down on GLUT4 trafficking and glucose uptake in L6 myoblasts. *PloS One*. 2013; 8(5): e65195. doi: 10.1371/journal.pone.0065195.
37. Antonescu CN, Diaz M, Femia G, Planas JV, Klip A. Clathrin-dependent and independent endocytosis of glucose transporter 4 (GLUT4) in myoblasts: regulation by mitochondrial uncoupling. *Traffic*. 2008; 9(7): 1173–90. doi: 10.1111/j.1600-0854.2008.00755.x.
38. Klop B, Elte JW, Cabezas MC. Dyslipidemia in obesity: mechanisms and potential targets. *Nutrients*. 2013; 5(4): 1218–40. doi: 10.3390/nu5041218.
39. Franssen R, Monajemi H, Stroes ES, Kastelein JJ. Obesity and dyslipidemia. *Med Clin North Am*. 2011; 95(5): 893–902. doi: 10.1016/j.mcna.2011.06.003.
40. Pohl J, Ring A, Ehehalt R, Schulze-Bergkamen H, Schad A, Verkade P, et al. Long-chain fatty acid uptake into adipocytes depends on lipid raft function. *Biochemistry*. 2004; 43(14): 4179–87. doi: 10.1021/bi035743m.
41. Pohl J, Ring A, Stremmel W. Uptake of long-chain fatty acids in HepG2 cells involves caveolae: analysis of a novel pathway. *J Lipid Res*. 2002; 43(9): 1390–9.
42. Ost A, Ortegren U, Gustavsson J, Nystrom FH, Stralfors P. Triacylglycerol is synthesized in a specific subclass of caveolae in primary adipocytes. *J Biol Chem*. 2005; 280(1): 5–8. doi: 10.1074/jbc.C400429200.
43. Uittenbogaard A, Everson WV, Matveev SV, Smart EJ. Cholestryler ester is transported from caveolae to internal membranes as part of a caveolin-annexin II lipid–protein complex. *J Biol Chem*. 2002; 277(7): 4925–31. doi: 10.1074/jbc.M109278200.
44. Katheria AC, Masliah E, Benirschke K, Jones KL, Kim JH. Idiopathic persistent pulmonary hypertension in an infant with Smith-Lemli-Opitz syndrome. *Fetal Pediatr Pathol*. 2010; 29(6): 373–9. doi: 10.3109/15513815.2010.512045.
45. Meshulam T, Breen MR, Liu L, Parton RG, Pilch PF. Caveolins/caveolae protect adipocytes from fatty acid-mediated lipotoxicity. *J Lipid Res*. 2011; 52(8): 1526–32. doi: 10.1194/jlr.M015628.
46. Howitt L, Grayson TH, Morris MJ, Sandow SL, Murphy TV. Dietary obesity increases NO and inhibits BKCa-mediated, endothelium-dependent dilation in rat cremaster muscle artery: association with caveolins and caveolae. *Am J Physiol Heart Circ Physiol*. 2012; 302(12): H2464–76. doi: 10.1152/ajpheart.00965.2011.
47. Su X, Abumrad NA. Cellular fatty acid uptake: a pathway under construction. *Trends Endocrinol Metab*. 2009; 20(2): 72–7. doi: 10.1016/j.tem.2008.11.001.
48. Grayson TH, Chadha PS, Bertrand PP, Chen H, Morris MJ, Senadheera S, et al. Increased caveolae density and caveolin-1 expression accompany impaired NO-mediated vasorelaxation in diet-induced obesity. *Histochem Cell Biol*. 2013; 139(2): 309–21. doi: 10.1007/s00418-012-1032-2.
49. Knowles CJ, Cebova M, Pinz IM. Palmitate diet-induced loss of cardiac caveolin-3: a novel mechanism for lipid-induced contractile dysfunction. *PloS One*. 2013;8(4):e61369. doi: 10.1371/journal.pone.0061369.
50. Mitsutake S, Zama K, Yokota H, Yoshida T, Tanaka M, Mitsui M, et al. Dynamic modification of sphingomyelin in lipid microdomains controls development of obesity, fatty liver, and type 2 diabetes. *J Biol Chem*. 2011; 286(32): 28544–55. doi: 10.1074/jbc.M111.255646.
51. Razani B, Combs TP, Wang XB, Frank PG, Park DS, Russell RG, et al. Caveolin-1-deficient mice are lean, resistant to diet-induced obesity, and show hypertriglyceridemia with adipocyte abnormalities. *J Biol Chem*. 2002; 277(10): 8635–47. doi: 10.1074/jbc.M110970200.
52. Takai M, Tsuzuki S, Matsuno Y, Kozai Y, Eguchi A, Matsumura S, et al. Assessment of key amino-acid residues of CD36 in specific binding interaction with an oxidized low-density lipoprotein. *Biosci Biotechnol Biochem*. 2013; 77(5): 1134–7.
53. Calvo D, Gomez-Coronado D, Suarez Y, Lasuncion MA, Vega MA. Human CD36 is a high affinity receptor for the native lipoproteins HDL, LDL, and VLDL. *J Lipid Res*. 1998; 39(4): 777–88.
54. Podrez EA, Poliakov E, Shen Z, Zhang R, Deng Y, Sun M, et al. Identification of a novel family of oxidized phospholipids that serve as ligands for the macrophage scavenger receptor CD36. *J Biol Chem*. 2002; 277(41): 38503–16. doi: 10.1074/jbc.M203318200.
55. Inagaki H, Tsuzuki S, Iino T, Inoue K, Fushiki T. Development of an in vitro system for screening the ligands of a membrane glycoprotein CD36. *Cytotechnology*. 2008; 57(2): 145–50. doi: 10.1007/s10616-008-9123-6.
56. Covey SD, Brunet RH, Gandhi SG, McFarlane N, Boreham DR, Gerber GE, et al. Cholesterol depletion inhibits fatty acid uptake without affecting CD36 or caveolin-1 distribution

- in adipocytes. *Biochem Biophys Res Commun.* 2007; 355(1): 67–71. doi: 10.1016/j.bbrc.2007.01.135.
57. Levitan I, Shentu TP. Impact of oxLDL on cholesterol-rich membrane rafts. *J Lipids.* 2011; 2011: 730209. doi: 10.1155/2011/730209.
58. Truong TQ, Brodeur MR, Falstrault L, Rhainds D, Brissette L. Expression of caveolin-1 in hepatic cells increases oxidized LDL uptake and preserves the expression of lipoprotein receptors. *J Cell Biochem.* 2009; 108(4): 906–15. doi: 10.1002/jcb.22321.
59. Sun B, Boyanovsky BB, Connelly MA, Shridas P, van der Westhuyzen DR, Webb NR. Distinct mechanisms for OxLDL uptake and cellular trafficking by class B scavenger receptors CD36 and SR-BI. *J Lipid Res.* 2007; 48(12): 2560–70. doi: 10.1194/jlr.M700163-JLR200.
60. Shaul PW. Endothelial nitric oxide synthase, caveolae and the development of atherosclerosis. *J Physiol.* 2003; 547(Pt 1): 21–33. doi: 10.1113/jphysiol.2002.031534.
61. Mineo C, Shaul PW. Regulation of eNOS in caveolae. *Adv Exp Med Biol.* 2012; 729: 51–62. doi: 10.1007/978-1-4614-1222-9_4.
62. Zhang WZ, Venardos K, Finch S, Kaye DM. Detrimental effect of oxidized LDL on endothelial arginine metabolism and transportation. *Int J Biochem Cell Biol.* 2008; 40(5): 920–8. doi: 10.1016/j.biocel.2007.10.027.
63. Ramirez-Sanchez I, Mendoza-Lorenzo P, Zentella-Dehesa A, Mendez-Bolaina E, Lara-Padilla E, Ceballos-Reyes G, et al. Caveolae and non-caveolae lipid raft microdomains of human umbilical vein endothelial cells contain utrophin-associated protein complexes. *Biochimie.* 2012; 94(9): 1884–90. doi: 10.1016/j.biochi.2012.05.001.
64. Dessy C, Feron O, Balligand JL. The regulation of endothelial nitric oxide synthase by caveolin: a paradigm validated in vivo and shared by the ‘endothelium-derived hyperpolarizing factor’. *Pflugers Arch.* 2010; 459(6): 817–27. doi: 10.1007/s00424-010-0815-3.
65. Sun J, Kohr MJ, Nguyen T, Aponte AM, Connelly PS, Esfahani SG, et al. Disruption of caveolae blocks ischemic preconditioning-mediated S-nitrosylation of mitochondrial proteins. *Antioxid Redox Signal.* 2012; 16(1): 45–56. doi: 10.1089/ars.2010.3844.
66. Bernatchez P, Sharma A, Bauer PM, Marin E, Sessa WC. A noninhibitory mutant of the caveolin-1 scaffolding domain enhances eNOS-derived NO synthesis and vasodilation in mice. *J Clin Invest.* 2011; 121(9): 3747–55. doi: 10.1172/JCI44778.
67. Lobysheva I, Rath G, Sekkali B, Bouzin C, Feron O, Gallez B, et al. Moderate caveolin-1 downregulation prevents NADPH oxidase-dependent endothelial nitric oxide synthase uncoupling by angiotensin II in endothelial cells. *Arterioscler Thromb Vasc Biol.* 2011; 31(9): 2098–105. doi: 10.1161/ATVBAHA.111.230623.
68. Shu L, Park JL, Byun J, Pennathur S, Kollmeyer J, Shayman JA. Decreased nitric oxide bioavailability in a mouse model of Fabry disease. *J Am Soc Nephrol.* 2009; 20(9): 1975–85. doi: 10.1681/ASN.2008111190.
69. Makarewich CA, Correll RN, Gao H, Zhang H, Yang B, Berretta RM, et al. A caveolae-targeted L-type Ca(2)+ channel antagonist inhibits hypertrophic signaling without reducing cardiac contractility. *Circ Res.* 2012; 110(5): 669–74. doi: 10.1161/CIRCRESAHA.111.264028.
70. Quintas LE, Pierre SV, Liu L, Bai Y, Liu X, Xie ZJ. Alterations of Na+ /K+ -ATPase function in caveolin-1 knockout cardiac fibroblasts. *J Mol Cell Cardiol.* 2010; 49(3): 525–31. doi: 10.1016/j.yjmcc.2010.04.015.
71. Linder AE, Thakali KM, Thompson JM, Watts SW, Webb RC, Leite R. Methyl-beta-cyclodextrin prevents angiotensin II-induced tachyphylactic contractile responses in rat aorta. *J Pharmacol Exp Ther.* 2007; 323(1): 78–84. doi: 10.1124/jpet.107.123463.
72. Sones WR, Davis AJ, Leblanc N, Greenwood IA. Cholesterol depletion alters amplitude and pharmacology of vascular calcium-activated chloride channels. *Cardiovasc Res.* 2010; 87(3): 476–84. doi: 10.1093/cvr/cvq057.
73. Morris JB, Huynh H, Vasilevski O, Woodcock EA. Alpha1-adrenergic receptor signaling is localized to caveolae in neonatal rat cardiomyocytes. *J Mol Cell Cardiol.* 2006; 41(1): 17–25. doi: 10.1016/j.yjmcc.2006.03.011.
74. Chai Q, Wang XL, Zeldin DC, Lee HC. Role of caveolae in shear stress-mediated endothelium-dependent dilation in coronary arteries. *Cardiovas Res.* 2013; 100(1): 151–9. doi: 10.1093/cvr/cvt157.
75. Gagnon C, Lu ZX, Magliano DJ, Dunstan DW, Shaw JE, Zimmet PZ, et al. Low serum 25-hydroxyvitamin D is associated with increased risk of the development of the metabolic syndrome at five years: results from a national, population-based prospective study (The Australian Diabetes, Obesity and Lifestyle Study: AusDiab). *J Clin Endocrinol Metab.* 2012; 97(6): 1953–61. doi: 10.1210/jc.2011-3187.
76. Boyan BD, Chen J, Schwartz Z. Mechanism of Pdia3-dependent 1alpha,25-dihydroxy vitamin D3 signaling in musculoskeletal cells. *Steroids.* 2012; 77(10): 892–6. doi: 10.1016/j.steroids.2012.04.018.
77. Haussler MR, Jurutka PW, Mizwicki M, Norman AW. Vitamin D receptor (VDR)-mediated actions of 1alpha,25(OH)(2)vitamin D(3): genomic and non-genomic mechanisms. *Best Pract Res Clin Endocrinol Metab.* 2011; 25(4): 543–59. doi: 10.1016/j.beem.2011.05.010.
78. Norman AW. Minireview: vitamin D receptor: new assignments for an already busy receptor. *Endocrinology.* 2006; 147(12): 5542–8. doi: 10.1210/en.2006-0946.
79. Buitrago C, Boland R. Caveolae and caveolin-1 are implicated in 1alpha,25(OH)2-vitamin D3-dependent modulation of Src, MAPK cascades and VDR localization in skeletal muscle cells. *J Steroid Biochem Mol Biol.* 2010; 121(1–2): 169–75. doi: 10.1016/j.jsbmb.2010.03.002.
80. Ren G, Jacob RF, Kaulin Y, Dimuzio P, Xie Y, Mason RP, et al. Alterations in membrane caveolae and BKCa channel activity in skin fibroblasts in Smith-Lemli-Opitz syndrome. *Mol Genet Metab.* 2011; 104(3): 346–55. doi: 10.1016/j.ymgme.2011.04.019.
81. Bowers LW, Cavazos DA, Brenner AJ, Hursting SD, Maximo IX, Degraffenreid LA. Obesity enhances nongenomic estrogen receptor crosstalk with the PI3K/Akt and MAPK pathways to promote in vitro measures of breast cancer progression. *Breast Cancer Res.* 2013; 15(4): R59. doi: 10.1186/bcr3453.
82. Watson CS, Jeng YJ, Hu G, Wozniak A, Bulayeva N, Guptarak J. Estrogen- and xenoestrogen-induced ERK signaling in pituitary tumor cells involves estrogen receptor-alpha interactions with G protein-alphai and caveolin I. *Steroids.* 2012; 77(5): 424–32. doi: 10.1016/j.steroids.2011.12.025.
83. Handgraaf S, Riant E, Fabre A, Waget A, Burcelin R, Liere P, et al. Prevention of obesity and insulin resistance by estrogens requires ERalpha activation function-2 (ERalphaAF-2), whereas ERalphaAF-1 is dispensable. *Diabetes.* 2013; 62(12): 4098–108. doi: 10.2337/db13-0282.
84. Matic M, Bryzgalova G, Gao H, Antonson P, Humire P, Omoto Y, et al. Estrogen signalling and the metabolic syndrome: targeting the hepatic estrogen receptor alpha action. *PloS One.* 2013;8(2):e57458. doi: 10.1371/journal.pone.0057458.
85. Finan B, Yang B, Ottaway N, Stemmer K, Muller TD, Yi CX, et al. Targeted estrogen delivery reverses the metabolic syndrome. *Nat Med.* 2012; 18(12): 1847–56. doi: 10.1038/nm.3009.
86. Manrique C, Lastra G, Habibi J, Mugerfeld I, Garro M, Sowers JR. Loss of estrogen receptor alpha signaling leads to

- insulin resistance and obesity in young and adult female mice. *Cardiorenal Med.* 2012; 2(3): 200–10. doi: 10.1159/000339563.
87. Fuentes E, Fuentes F, Vilahur G, Badimon L, Palomo I. Mechanisms of chronic state of inflammation as mediators that link obese adipose tissue and metabolic syndrome. *Mediators Inflamm.* 2013; 2013: 136584. doi: 10.1155/2013/136584.
88. Simmons GE, Jr., Taylor HE, Hildreth JE. Caveolin-1 suppresses human immunodeficiency virus-1 replication by inhibiting acetylation of NF-kappaB. *Virology.* 2012; 432(1): 110–19. doi: 10.1016/j.virol.2012.05.016.
89. Di Raimondo D, Tuttolomondo A, Butta C, Casuccio A, Giarrusso L, Miceli G, et al. Metabolic and anti-inflammatory effects of a home-based programme of aerobic physical exercise. *Int J Clin Practice.* 2013; 67(12): 1247–53. doi: 10.1111/ijcp.12269.
90. Grundy SM, Cleeman JI, Daniels SR, Donato KA, Eckel RH, Franklin BA, et al. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. *Circulation.* 2005; 112(17): 2735–52. doi: 10.1161/CIRCULATIONAHA.105.169404.
91. Suliburska J, Bogdanski P, Szulinska M, Pupek-Musialik D, Jablecka A. Changes in mineral status are associated with improvements in insulin sensitivity in obese patients following L-arginine supplementation. *Eur J Nutr.* 2013. doi: 10.1007/s00394-013-0533-7
92. Poyrazoglu S, Bas F, Darendeliler F. Metabolic syndrome in young people. *Curr Opin Endocrinol Diabetes Obes.* 2013; 21(1): 56–63. doi: 10.1097/01.med.0000436414.90240.2c.